Note

Structural studies of the O-specific side-chains of the lipopolysaccharide from Yersinia enterocolitica Ye 128

JAMES HOFFMAN, BENGT LINDBERG,

Department of Organic Chemistry, Arrhenius Laboratory, University of Stockholm, S-106 91 Stockholm (Sweden)

AND ROBERT R. BRUBAKER

Department of Microbiology and Public Health, Michigan State University, East Lansing, Michigan 48824 (U.S.A.)

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The presence of the unusual sugar 6-deoxy-L-altrose as a component of the lipopolysaccharide (LPS) from a strain of *Yersinia enterocolitica* has been reported¹. Sugar analyses of LPS from strains of this organism indicate that this sugar is present in LPS from different serotypes^{2,3}. We now report studies of the LPS from *Y. enterocolitica* Ye 128 (Daniels 924) belonging to serotype 2 of Winblad⁴ or serogroup IB of Knapp and Thal⁵.

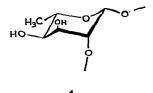
The polysaccharide (PS) was prepared from the LPS by acid hydrolysis under mild conditions. An acid hydrolysate of the PS contained a sugar which was chromatographically indistinguishable from 6-deoxy-D-altrose⁶, together with traces of glucose, heptose, and another minor component, tentatively identified as rhamnose. The major component was isolated as a chromatographically pure syrup. Its ¹H-n.m.r. spectrum and that given by authentic 6-deoxy-D-altrose were superposable. The natural sugar has $[\alpha]_{578}$ —14° (c 2, water), compared to +16° for the D sugar⁷, demonstrating that the former has the L configuration.

Methylation analysis of the PS gave a single, major component, identified from the mass spectrum of its alditol acetate as 6-deoxy-3,4-di-O-methyl-L-altrose. Consequently, the sugar residues in the PS are pyranosidic and linked through the 2-position.

The PS had $[\alpha]_{578}$ +72° (c 1, water), indicating that the 6-deoxy-L-altropyranosyl residues have the β configuration. In agreement with this conclusion, the signal for the anomeric proton in the ¹H-n.m.r. spectrum appeared at δ 5.16 ($J_{1,2}$ low), compared to the value δ 5.09 ($J_{1,2}$ 1.3 Hz) reported for β -D-altropyranose⁸. The presence of only six strong signals in the ¹³C-n.m.r. spectrum confirmed that the major component of the PS was the O-antigenic chain, having a simple structure. The signal for C-1 appeared at 101.3 p.p.m. with a $J_{13C,H}$ value of 164 Hz, demonstrating that H-1 is axial⁹. The combined evidence therefore demonstrates that the

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O-antigen is a homopolymer composed of $(1\rightarrow 2)$ -linked 6-deoxy- β -L-altropyranosyl residues, and that these residues are in the ${}^{1}C_{4}$ conformation, as in 1.



EXPERIMENTAL

General methods. — Concentrations were performed under reduced pressure at bath temperatures not exceeding 40°. G.l.c. was performed with a Perkin-Elmer model 990 instrument fitted with flame-ionisation detectors. Separations were performed on 3% of OV-225 on Gas Chrom W (100-200 mesh) in glass columns (180 × 0.15 cm). G.l.c.-m.s. was performed on a Varian Mat 311 instrument fitted with an OV-225 column. For n.m.r. spectra, a Jeol FX 100 instrument was used. The spectra were obtained for solutions in D_2O at 85°, using external tetramethylsilane (^{13}C -n.m.r.) or internal sodium 1,1,2,2,3,3-hexadeuterio-4,4-dimethyl-4-silapentane-1-sulfonate (14 -n.m.r.) as references. Undecoupled spectra were obtained by the gated decoupling technique with a sampling time of 0.4 sec and a pulse repetition time of 1 sec. Optical rotations were determined with a Perkin-Elmer 241 instrument. Methylation was performed by the Hakomori method with sodium methylsulfinylmethanide-methyl iodide in dimethyl sulfoxide. Hydrolysis and analysis of the products were performed as previously described 11.

Identification of sugar component. — Acid hydrolysis of the polysaccharide gave one major component, identified by g.l.c.-m.s. of its alditol acetate as a 6-deoxy-hexose. This sugar was separated by p.c. on Whatman No. 1 paper, using 1-butanol-pyridine-water (6:4:3) as irrigant.

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REFERENCES

- 1 D. C. ELLWOOD AND G. R. A. KIRK, Biochem. J., 122 (1971) 14P.
- 2 K. WARTENBERG, J. LYSY, AND W. KNAPP, Zentralbl. Bakteriol. Parasitenk. Infektionskr. Hyg. Abt. 1: Orig. Reihe A, 230 (1975) 361-366.
- 3 W. Beer and G. Seltmann, Z. Allg. Mikrobiol., 13 (1973) 167-169.
- 4 S. WINBLAD, Symp. Ser. Immunobiol. Stand., 9 (1968) 337-342.

- 5 W. Knapp and E. Thal, Zentralbl. Bakteriol. Parasitenk. Infektionskr. Hyg. Abt. 1: Orig. Reihe A, 223 (1973) 88-105.
- 6 H. KAUFMANN, P. MÜHLRADT, AND T. REICHSTEIN, Helv. Chim. Acta, 50 (1967) 2287-2298.
- 7 M. GUT AND D. A. PRINS, Helv. Chim. Acta, 29 (1946) 1555-1559.
- 8 S. J. ANGYAL AND V. A. PICKLES, Aust. J. Chem., 25 (1972) 1695-1710.
- 9 K. BOCK, J. LUNDT, AND C. PEDERSEN, Tetrahedron Lett., (1973) 1037-1040.
- 10 S. HAKOMORI, J. Biochem. (Tokyo), 55 (1964) 205-208.
- 11 P. E. Jansson, L. Kenne, H. Liedgren, B. Lindberg, and J. Lönngren, Chem. Commun. Univ. Stockholm, 8 (1976).